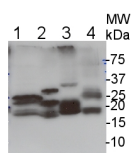


Product no **AS08 282****Lhc1 | from PSI of red alga****Product information**

Immunogen	Purified LHC complex from <i>Porphyridium cruentum</i>
Host	Rabbit
Clonality	Polyclonal
Purity	Serum
Format	Lyophilized
Quantity	200 µl
Reconstitution	For reconstitution add 200 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution	1 : 1000 (WB)
Expected apparent MW	20-25 kDa
Confirmed reactivity	<i>Aureococcus anophagefferens</i> , <i>Guillardia theta</i> , <i>Heterosigma akashiwo</i> , <i>Thalassiosira pseudonana</i> , <i>Porphyridium cruentum</i>
Predicted reactivity	Diatoms and other heterokonts, cryptophyte algae, red algae Species of your interest not listed? Contact us
Not reactive in	Higher plants
Additional information	Strongly reactive to 7 Lhc1 light harvesting polypeptides of <i>P. cruentum</i>
Selected references	Tan et al. (1995) . Decrease of polypeptides in the PS I antenna complex with increasing growth irradiance in the red alga <i>Porphyridium cruentum</i> . <i>Photosyn. Research</i> 45:1. Wolfe et al. (1994) Evidence for a common origin of chloroplasts with light-harvesting complexes of different pigmentation. <i>Nature</i> 367:566

Application example

2 µg of total chlorophyll/lane of total cell extract from (1) *Thalassiosira pseudonana*, (2) *Heterosigma akashiwo*, (3) *Guillardia theta*, (4) *Aureococcus anophagefferens*, extracted with Agrisera protein extraction buffer [PEB](#), were separated on 13-17% SDS-PAGE and blotted 2h to nitrocellulose. membranes were blocked 1h with 5% low-fat milk powder in PBS-T (0.1% TWEEN 20) and probed with anti-Lhc1 (AS08 282, 1:6000, 1h) and secondary anti-rabbit (1:5000, 1h) antibody (HRP conjugated) in PBS-T containing 2% low fat milk powder. Antibody incubations were followed by washings in PBS-T (15, +5, +5, +5 min). All steps were performed at RT with agitation. Signal was detected with chemiluminescent detection reagent. Exposure time was 2 min.

Courtesy of Meriem Alami and Beverley Green, Canada