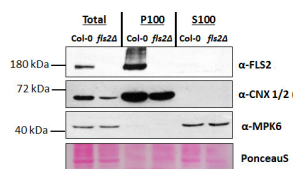


Product no **AS12 1857****FLS2 | Flagellin-sensitive 2****Product information**

Immunogen	KLH-conjugated peptide derived from <i>Arabidopsis thaliana</i> FLS2 sequence, UniProt: Q9FL28 , TAIR: AT5G46330
Host	Rabbit
Clonality	Polyclonal
Purity	Immunogen affinity purified serum in PBS pH 7.4.
Format	Lyophilized
Quantity	50 µg
Reconstitution	For reconstitution add 50 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles, Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution	1 : 5000 (WB)
Expected apparent MW	126 kDa (without propeptide) 175-180 kDa
Confirmed reactivity	<i>Arabidopsis thaliana</i>
Predicted reactivity	<i>Capsella rubella</i> , <i>Glycine max</i> , <i>Hordeum vulgare</i> , <i>Populus trichocarpa</i> , <i>Solanum lycopersivum</i> , <i>Vitis vinifera</i> Species of your interest not listed? Contact us
Not reactive in	No confirmed exceptions from predicted reactivity are currently known
Selected references	Kalischuk et al. (2022) Amplification of cell signaling and disease resistance by an immunity receptor Ve1Ve2 heterocomplex in plants. <i>Commun Biol.</i> 2022 May 25;5(1):497. doi: 10.1038/s42003-022-03439-0. PMID: 35614138; PMCID: PMC9132969. Wang et al. (2021) Arabidopsis PUB2 and PUB4 connect signaling components of pattern-triggered immunity. <i>New Phytol.</i> 2021 Dec 17. doi: 10.1111/nph.17922. Epub ahead of print. PMID: 34918346. Ngou et al. (2021) Mutual potentiation of plant immunity by cell-surface and intracellular receptors. <i>Nature.</i> 2021 Mar 10. doi: 10.1038/s41586-021-03315-7. Epub ahead of print. PMID: 33692545. Hilleary et al. (2020) . Tonoplast-localized Ca ²⁺ pumps regulate Ca ²⁺ signals during pattern-triggered immunity in <i>Arabidopsis thaliana</i> . <i>Proc Natl Acad Sci U S A.</i> 2020 Aug 4;117(31):18849-18857. doi: 10.1073/pnas.2004183117. Zhang et al. (2019) . An important role of L-fucose biosynthesis and protein fucosylation genes in <i>Arabidopsis</i> immunity. <i>New Phytol.</i> 2018 Dec 15. doi: 10.1111/nph.15639.

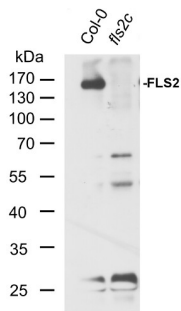
Application example

Proteins were isolated from 7-day old *Arabidopsis thaliana* seedlings, Col-0 (wild-type) and fls2 null mutant (SALK_093905; Heese et al., 2007). Total proteins (T) were isolated and then fractionated by ultracentrifugation at 100,000 x gravity for 30 min at 4°C into soluble (S100) and microsomal (P100) proteins as described in LaMontagne et al. (2016). For each fraction, 30 µg of proteins were denatured at 65°C for 5 min, separated on an 8% SDS-PAGE and transferred for 1h using a tank transfer system to nitrocellulose membrane. Blots were blocked with 1x PBS (Fisher Scientific BP665-1) + 0.1% Tween 20 (PBS-T) + 5% milk for 1.5h at room temperature (RT) with agitation. Blots were incubated with primary antibody at a dilution of 1:1000 overnight at 4°C with agitation in 1x PBS-T + 5% milk. The antibody solution was decanted, and the blot washed four times (1x5min and 3x6min) in 1x PBS-T at RT with agitation. Blot was incubated with secondary antibody [Goat anti Rabbit IgG (H&L) –HRP conjugated; [AS09 602](#)] diluted to 1:10 000 in 1x PBS-T + 5% milk for 3h 15min at RT with agitation. The blot was washed as above and developed for 5 min with chemiluminescent detection reagent. Exposure time was 5min. -CNX 1/2 ([AS12 2365](#), Agrisera) and -MPK6 (Heese et al., 2007) confirmed separation of microsomal from soluble fraction.

Courtesy of Gayani Ekanayke¹, Grant Mc Gowan^{1,2} & Dr. Antje Heese¹ [1 Div. of Biochemistry, Interdisciplinary Plant Group (IPG) - University of Missouri; Columbia, MO, 65211, USA; 2 Dep. of Molecular & Cell Biology, University of Cape Town, Rondebosch 7701, South Africa].

References [Heese, A. et al. \(2007\)](#). The receptor-like kinase SERK3/BAK1 is a central regulator of innate immunity in plants. *PNAS*. Vol. 104 no. 29 p. 12217–12222.

[LaMontagne, E. et al. \(2016\)](#). Isolation of Microsomal Membrane Proteins from *Arabidopsis thaliana*. *Current Protocols in Plant Biology* 1:1-18. doi: 10.1002/cppb.20020.



Plant material: *Arabidopsis thaliana* Col-0 (wild type) fls2c mutant (described in [Nekrasov et al. 2009](#)). Method: Two-week-old seedlings (grown under short day conditions, 8h light) were collected and frozen in liquid nitrogen. Approximately 100mg of plant material were extracted in 0.2 ml of homogenization buffer (250 mM sucrose, 50 mM HEPES-KOH pH 7.5, 5% glycerol, 0.5% Triton X-100, 50 mM Na₄P₂O₇, 1 mM Na₂MoO₄, 25 mM NaF, 2 mM DTT, Sigma plant protease inhibitor cocktail) with a glass pistil and a small amount of sand in an 1.5ml Eppendorf tube. Another 0.8 ml of buffer were added and the extract was mixed thoroughly. Debris was pelleted by centrifugation in a table-top microcentrifuge at 13 000 rpm. The supernatant was mixed with 4x SDS loading buffer (200 mM TRIS-HCl pH 6.8, 400 mM DTT, 8% SDS, 40% glycerol, 0.1% bromophenol blue) and boiled at 95 °C for 5min. 10µl were run on an 10% polyacrylamide gel and blotted onto a 0.45µm PVDF membrane (Carl Roth). The membrane was blocked for 1h in TBS-T (150mM NaCl, 10mM Tris-HCl pH8, 0.05% Tween-20) containing 5% skimmed milk powder. The primary antibody (Agrisera Rabbit anti-FLS2, 1µg/µl) was diluted 1:5000 in TBS-T containing 5% milk powder and incubated on the membrane overnight at 4 °C. Then the membrane was washed 5 times 15min with TBS-T containing 5% milk powder. The secondary antibody was Agrisera goat-anti rabbit IgG HRP-conjugated ([AS09 602](#)) at a dilution of 1:5000 diluted in TBS-T containing 5% milk powder in the same solution and incubated on the membrane at room temperature for 2h. The membrane was then washed 5 times 15min with TBS-T (no milk powder) and the blot was developed using chemiluminescent detection reagent.

Courtesy of Dr. Elena Petutsching, Georg-August-University Goettingen, Germany